

## Product Overview

This is 2X concentrated solution for PCR reaction which contains dNTPs, Taq DNA Polymerase, Mg<sup>2+</sup> and other critical reaction components. It doesn't contain primers and DNA template. Leo Mastermix is designed for routine PCR and also for amplification of high GC templates.

## Catalog Details

|       |                |
|-------|----------------|
| R8120 | 100 Reactions  |
| R8121 | 1000 Reactions |
| R8122 | 4000 Reactions |

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## Storage

- 20 °C

## PCR Protocol

The samples must be prepared in sterile DNase Free micro-centrifuge tubes, with the following composition placed on ice

| Components            | Example for 50µL reaction | Final Concentration |
|-----------------------|---------------------------|---------------------|
| Template DNA          | 1 µL                      | 5ng - 100 ng        |
| Forward Primer (10µM) | 1 µL                      | 0.1 - 1µM           |
| Reverse Primer (10µM) | 1 µL                      | 0.1 - 1µM           |
| Leo MasterMix (2X)    | 25 µL                     | 1X                  |
| Nuclease Free Water   | Upto 50 µL                |                     |

### NOTES:

- To collect all liquid at the bottom of the vial, reaction mix can be kept for a quick spin (10 seconds).
- While doing PCR without heat lid, it is recommended to overlay the sample with 1-2 drops of mineral oil.

After the preparation of PCR mix samples as above, they are transferred to thermal cycler. The thermal cycling process begins with initial denaturation step. The denaturation process is to ensure complete denaturation of the target DNA at 95°C. The standard steps for thermal cycler are tabulated below with optimum temperature, time, and number of cycles. Generally, 25 – 35 cycles yield sufficient product.

| Step                 | Temperature | Time            | Cycle   |
|----------------------|-------------|-----------------|---------|
| Initial denaturation | 95 ± 1 °C   | 2 - 5 minutes   | 1       |
| Denaturation         | 95 ± 1 °C   | 0.5 - 1 minutes | 25 - 35 |
| Annealing *          | 45 - 65 °C  | 0.5 - 1 minutes |         |
| Extension            | 68 - 72 °C  | 1 minute/kb     | 1       |
| Final Extension      | 68 - 72 °C  | 5 - 15 minutes  |         |
| Hold, if required.   | 2- 8 °C     | variable        | 1       |

If your mastermix contains green dye, then load it directly on the gel. The green dye will separate into blue (~4kb) and yellow (~25bp) in 1% agarose gel.

## COLONY PCR METHOD



1. Take a pipette tip (10 $\mu$ L) and just touch the colony. Add this colony to LEO mastermix.
2. Follow the PCR program as suggested or your own. Initial denaturation step must be 95°C for 2 minutes.
3. After PCR, directly load the sample to gel. YOU ARE DONE.

### NOTE:

- No need to add entire colony.
- No need to add even half the colony.
- Just a touch is all required.

## Quality Control Assays

1. **Purity:** SDS Page analysis with Coomassie Blue Staining resulted in  $\geq 95\%$  purity.
2. **Performance testing:** In a 25 $\mu$ L reaction, 12.5 $\mu$ L of mastermix was used to amplify 1ng of DNA template ( $\lambda$ ) of various sizes (300bp, 1kb, 3kb, 5kb) with appropriate primers. PCR was run with 30 cycles resulted in a single band, confirmed by 1% agarose gel electrophoresis with EtBr.
3. **Nuclease tests:** No contamination of endo or exonucleases were detected.

## Any Technical Help ?

Please write to us at [tech@dxbidt.com](mailto:tech@dxbidt.com) . Response can be expected within 24Hrs. Our technical team shall be happy to assist you all the time.

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